Pergamon Pergamon

0957-4166(94)00170-7

Asymmetric Synthesis of the Natural Diastereoisomer of trans-α-(2-Carboxymethylcyclopropyl)glycine isolated from Blighia unijugata

Carmen Alcaraz and Manuel Bernabé*

Instituto de Química Orgánica, CSIC, Juan de la Cierva 3, 28006-Madrid (Spain)

Abstract: An asymmetric synthesis of $(2S, 1.S, 2.R)-\alpha-(2\text{-carboxymethylcyclopropyl})$ glycine, formerly isolated from *Blighia unijugata* is reported. The key step is the dibromocyclopropanation of the oxazolidine alkenes 2, prepared in five steps from *D*-serine, which takes place with good diastereoisometric ratios. The major dibromocyclopropane was reduced to the corresponding cyclopropane, which was transformed in a single step into the title cyclopropylglycine.

Recently a number of new amino acids have been discovered in nature.¹ Among them, those containing a cyclopropane ring have attracted special attention due to their biological activities as phytochemical agents, enzyme inhibitors or probes in metabolism studies.²

Cyclopropylglycines form the largest group of naturally occurring cyclopropane amino acids, but only a few papers have been published on their synthesis, mainly referring to the synthesis of α -(methylene-cyclopropyl)glycine and α -(2-carboxycyclopropyl)glycine. We have recently described the asymmetric synthesis of $(E)-\alpha$ -[2-phenyl(ethyl)cyclopropyl)glycines³¹ from serine by diastereoselective dibromocyclopropanation, and we report now on the asymmetric synthesis of $trans-\alpha$ -(2-carboxymethylcyclopropyl)glycine, a natural product first isolated from the seeds of *Blighia unijugata*, which is a homolog of $trans-\alpha$ -(2-carboxycyclopropyl)glycine, known as a NMDA receptor agonist.

The oxazolidine aldehyde 1 was easily prepared in four steps from D-Serine in 65% overall yield.⁵ Wittig reaction of this aldehyde with the ylide derived from 2-(1,3-dioxan-2-yl)ethylidenetriphenyl phosphonium bromide⁶ by treatment with sodium hydride, gave a 80:20 mixture of Z and E alkenes 2 (Scheme 1). The major alkene was assumed to have Z configuration, the preferred one in this reaction with non-stabilized vlides.⁷

The mixture of alkenes 2 was refluxed with N-bromosuccinimide in benzene to produce a 66:33 mixture of both alkenes, which could not be separated by chromatography.

Scheme 1. Reagents: (a) 2-(1,3-dioxan-2-yl)ethylidenetriphenylphosphonium bromide, 95% HNa, THF (argon), r.t., 2h.

- (b) CHBr₃, KOH (s), TEBA, benzene, t < 30 °C,))), 18h.
 - (c) n-Bu₃SnH, hexane (argon), reflux, 24h.
- (d) 1. CrO₃, H₂SO₄, acetone-H₂O, r.t., 3h. 2. Dowex 50W (H⁺ form)

Addition of dibromocarbene, generated by the phase-transfer catalytic technique, 8 to the mixture of alkenes 2 (Z/E = 2/1) gave, after 10-15 days, a mixture of two dibromocyclopropyl derivatives, both of them having trans-relative configuration. An isomerization of Z- to E-alkene took first place and dibromocarbene was exclusively added to the E-isomer in a good diastereoisomeric ratio (3a/3b = 4/1), although in poor yield (20%).

It has been described that reaction times and yields in the addition of dichlorocarbene to olefins when this is generated under sonication, compare favorably with analogous phase-transfer procedures. In our case, when the addition of dibromocarbene to alkenes 2 was carried out with efficient stirring and ultrasonic irradiation, using a large excess (10 equivalents) of powdered potassium hydroxide and bromoform and a catalytic amount of triethylbenzylammonium chloride, a mixture of dibromocyclopropanes was obtained in 73% yield. The use of the phase-transfer catalyst was not strictly necessary, but the addition was faster in its presence. Under these conditions a mixture of three bromides (3a:3b:4) in a 4:1:2 ratio was obtained. One of the dibromocyclopropanes, compound 4, possesses cis-relative configuration, in contrast with results obtained without sonication. The facial stereoselectivity in the dibromocyclopropanation of E-alkene is the same in both cases.

The two major compounds (3a/4 = 2/1) refused to be separated by chromatographic means hence the mixture of both was reduced with two molar equivalents of tributyltin hydride to give the corresponding cyclopropanes 5a and 6 (85%), which were easily separated by flash chromatography.

Structural analyses were carried out for compounds 5a and 6 on the basis of 1D and 2D ¹H-NMR spectroscopic data in order to determine their relative configurations. Thus, analysis of the ¹H-NMR spectra of compounds 5a and 6 using the PANIC program gave the relevant coupling constants shown in Table 1. The values of the coupling constants of the geminal protons H-6 and H-7 (see Scheme 1) with the cyclopropane protons H-4 and H-5 indicate a *trans* configuration for compound 5a and a *cis* configuration for 6. On the other hand, the large coupling constant of 8.4 Hz suggests a dihedral angle of about 170° between protons H-3 and H-4 in 5a.

Comp.	J _{1,2}	J _{1,3}	J _{2,3}	J _{3,4}	J _{4,5}	J _{4,6}	J _{4,7}	J _{5,6}	J _{5,7}	J _{6,7}
5a	-9.0	5.6	1.1	8.4	4.3	8.4	4.6	4.7	8.5	-4.9
6	-8.7	5.6	2.0	8.8	8.6	8.7	5.9	8.6	5.9	-4.8

Table 1. Relevant Vicinal Coupling Constants (3J, Hz) found for Compounds 5a and 6

The 2D-NOESY spectra of compound 5a show, in addition to the expected H-4/H-6, H-5/H-7, H-3/H-5 and H-3/H-7 crosspeaks, which strongly support the *trans* configuration of the cyclopropane ring, a crosspeak H-2/H-7 which, in conjunction with the value of J_{3,4}, suggests a configuration 1'S, 2'R for the cyclopropane moiety, as depicted in Scheme 1.

Compound 5a was treated with Jones' reagent to give, after ion exchange chromatography, 75% of the natural trans-2-(2'-carboxymethylcyclopropyl)glycine (7)¹⁰: $[\alpha]_D$ +13 (c 0.67, H₂O), lit.⁴ $[\alpha]_D$ +12 (c 1, H₂O).

A previous synthesis of compound 7, starting from L-serine, gave an absolute value of $[\alpha]_D$ equal to that of the natural compound but with opposite sign. Thus, we propose an absolute configuration 2S, 1'S, 2R for the natural diastereoisomer of compound 7. Unfortunately an authentic sample was not available.

Acknowledgements. We are indebted to the Comunidad de Madrid for financial support (Grant C126/91)

REFERENCES

- (a) Wagner, I.; Musso, H. Angew. Chem. Int. Ed. Engl. 1983, 22, 816-828. (b) Rosenthal, G.A. Plant Nonproteinogenic Amino and Imino Acids. Biological and Toxicological Properties, Academic Press, New York, 1982.
- For a review, see Suckling, C.J. Angew. Chem. Int. Ed. Engl. 1988, 27, 537-552 and references therein.
 (b) Arenal, I.; Bernabé, M.; Fernández-Alvarez, E.; Gibello, A.M. An. Quím. 1983, 79 C, 65-68. (c) Bernabé, M.; Cuevas, O.; Fernández-Alvarez, E. Ibid. 1979, 75, 977-984. (d) Pirrung, M.C.; McGeehan, C.M. J. Org. Chem. 1986, 51, 2103-2106.
- (a) Black, D.; Landor, S. J. Chem. Soc., C, 1968, 288-290. (b) Kato, K; Takita, T.; Umezawa, H. Tetrahedron Lett., 1980, 21, 4925-4926. (c) Lawson, P.; McCarthy, M.; Sargerson, A. J. Am. Chem. Soc., 1982, 104, 6710-6716. (d) Landor, S.; Landor, D.; Kalli, M. J. Chem. Soc. Perkin Trans I, 1983, 2921-2925. (e) Kurokawa, N.; Ohfune, Y. Tetrahedron Lett., 1985, 26, 83-84. (f) Yamanoi, K.; Ohfune, Y. Tetrahedron Lett., 1988, 29, 1181-1184. (g) Shimamoto, K.; Ohfune, Y. Tetrahedron Lett., 1989, 30, 3803-3804. (h) Pelliciari, R.; Natalini, B.; Marinozzi, M.; Monahan, J.; Snyder, J. Tetrahedron Lett., 1990, 31, 139-142. (i) Shimamoto, K.; Ohfune, Y. Tetrahedron Lett., 1990, 31, 4049-4052. (j) Wessjohan, L.; Krass, N.; Yu, D.; deMeijere, A. Chem. Ber., 1992, 125, 867-882. (k) Es-Sayed, M.; Gatkowski, C.; Krass, N.; Meyers, A.I.; deMeijere, A. Synlett, 1992, 962-964. (l) de Frutos, M.P.; Fernández, M.D.; Fernández-Alvarez, E.; Bernabé, M. Tetrahedron, 1992, 48, 1123-1130.
- 4. Fowden, L.; Mac Gibbon, C.M.; Mellon, F.A.; Sheppard, R.C. Phytochemistry, 1972, 11, 1105-1110.
- 5. Garner, P.; Park, J.M. J. Org. Chem., 1987, 52, 2361-2364.
- 6. Stowell, J.C.; Keith, D.R. Synthesis, 1979, 132-133.
- 7. Maryanoff, B.E.; Reitz, A.B. Chem. Rev., 1989, 89, 863-927.
- 8. Dehmlow, S.S. Phase Transfer Catalysis; Verlag Chemie, 1983, pp. 220-285.
- 9. Regen, S.L.; Singh, A. J. Org. Chem., 1982, 47, 1587-1588.
- 10. ¹H-NMR (D₂O, 30 °C) (analysed by the PANIC program): δ 3.159 (d, J_{1,2} = 9.9, H-1), 2.504 (dd, J_{3,7} = 6.3, J_{6,7} = -17.2, H-7), 2.241 (dd, J_{3,6} = 8.2, J_{6,7} = -17.2, H-6), 1,123 (m, J_{2,3} = 4.4, J_{3,4}= 8.6, J_{3,5} = 5.3, J_{3,6} = 8.2, J_{3,7} = 6.3, H-3), 1.103 (m, J_{1,2} = 9.9, J_{2,3} = 4.4, J_{2,4} = 5.2, J_{2,5} = 8.9, H-2), 0.930 (dt, J_{2,5} = 8.9, J_{3,5} = 5.3, J_{4,5} = -5.5, H-5), 0.753 (dt, J_{2,4} = 5.2, J_{3,4} = 8.6, J_{4,5} = -5.5, H-4).